

# *GeneGroupAnalysisBetaCS*: a package to perform gene-set significance analysis for data sets with cross-sectional or time series designs.

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# 1 Introduction

The *GeneGroupAnalysis* package provides functions to perform statistical identification of gene functional classes that behave in a distinct manner between the phenotypes of interest for data sets under cross-sectional or time series designs. This package includes (i) functions to perform gene set comparison (ii) examples to visualize the results of such comparisons.

## 1.1 Installation

*GeneGroupAnalysisBetaCS* requires that *MCMCpack*, *AnnotationDbi*, *annotate*, *tcltk* and *R* ( $\geq 2.10.0$ ) are installed. These should be installed automatically when you install *GeneGroupAnalysisBetaCS*. To install *GeneGroupAnalysis*, source *biocLite* from *bioconductor*:

```
> source("http://bioconductor.org/biocLite.R")
> biocLite("GeneGroupAnalysisBetaCS")
```

Load the *GeneGroupAnalysis*, into your current workspace:

```
> library(GeneGroupAnalysisBetaCS)
```

## 1.2 Further help

To view the *GeneGroupAnalysisBetaCS* description and a summary of all the functions within *GeneGroupAnalysisBetaCS*, type the following:

```
> library(help=GeneGroupAnalysisBetaCS)
```

## 1.3 Citing

We are delighted if you use this package. Please do email us if you find a bug or have a suggestion. We would be very grateful if you could cite:

Quiroz-Zarate A and Quackenbush J (2012). *Manuscript in preparation* .

# 2 Two simple cases: from gene expression to functional class selection.

We will very briefly demonstrate the use of some functions in *GeneGroupAnalysisBetaCS* by providing its application on two data sets with different experimental design.

We use the *breastCancerVDX* data library from *Bioconductor* for demonstration purposes under a cross-sectional design. This data set corresponds to the data set from [1].

Minn, AJ and colleagues used Affymetrix U133A Gene Chips to profile gene expression in 286 fresh-frozen tumor samples from patients with lymph-node-negative breast cancer who were treated during 1980 – 95, but who did not receive systemic neoadjuvant or adjuvant therapy. These samples correspond from the data set used in [3] with GEO reference accession number GSE2034, from the tumor bank at the Erasmus Medical Center in Rotterdam, Netherlands. An additional 58 estrogen receptor-negative samples were added from [1] GEO (GSE5327). In total 209 tumor samples are classified as ER+ and 135 as ER-. Even though this data set comes from a 5-year follow-up design, the way the data is conceived for this analysis is cross-sectional. For purposes of providing an example on the use of the functions on the package the data set with cross-sectional design has a 70% of the samples with ER+ and with ER- where selected randomly selected.

## 2.1 Example: Data analysis under a cross-sectional setting.

This is an example on how to perform an analysis with the proposed method in [2] for a data set with cross-sectional design. This example is divided in two parts. The data preparation and the execution of the Gibb's sampler function.

### 2.1.1 Data preprocessing stage

The original gene expression data set Minn AJ and colleagues [1] has a U133A Affymetrix platform. The normalized data set was saved to the variable `vdx` in the *breastCancerVDX* data library from Bioconductor.

```
> library(breastCancerVDX)
> library(GeneGroupAnalysisBetaCS)
> library(hgu133a.db)
> library(annotate)
> data(vdx,package="breastCancerVDX")
> #-Normalized expression data set
> minn.data.expr=exprs(vdx)
> #--- Checking that the columns correspond to their respective phenotype data id
> #all(colnames(minn.data.expr)==rownames(pData(vdx)))
```

The data on `minn.data.expr` has the following appearance (maybe with a different column order):

	ER +			ER -		
	Sample 1	...	Sample 209	Sample 1	...	Sample 135
Probe ID 1	7.84	...	7.31	7.11	...	6.98
⋮	⋮	⋮	⋮	⋮	⋮	⋮
Probe ID <i>q</i>	6.14	...	6.81	6.52	...	6.37

The rows correspond to the measurements for the Affymetrix identifiers and the columns correspond to the gene expression measurements on the samples.

### 3 Session Info

- R version 2.15.0 (2012-03-30), x86\_64-apple-darwin9.8.0
- Locale: C/en\_US.UTF-8/en\_US.UTF-8/C/en\_US.UTF-8/en\_US.UTF-8
- Base packages: base, datasets, grDevices, graphics, methods, stats, utils
- Other packages: AnnotationDbi 1.18.0, Biobase 2.16.0, BiocGenerics 0.2.0, DBI 0.2-5, GeneGroupAnalysisBetaCS 1.0, MASS 7.3-18, MCMCpack 1.2-3, RSQLite 0.11.1, annotate 1.34.0, breastCancerVDX 1.0.3, coda 0.14-7, hgu133a.db 2.7.1, lattice 0.20-6, org.Hs.eg.db 2.7.1
- Loaded via a namespace (and not attached): IRanges 1.14.2, grid 2.15.0, stats4 2.15.0, tcltk 2.15.0, tools 2.15.0, xtable 1.7-0

### References

- [1] Minn AJ, Gupta GP, Padua D, Bos P, Nguyen DX, Nuyten D, Kreike B, Zhang Y, Wang Y, Ishwaran H, Foekens JA, Van de Vijver M and Massagué J: Lung Metastasis Genes Couple Breast Tumor Size and Metastatic Spread. *PNAS*, **104(16)**, 6740-6745. 2007.
- [2] Quiroz-Zarate A and Quackenbush J XXXX: Genes as repeated measures of gene-set significance *Journal* **Vol(Num):Page 1-Page N**. 2011.
- [3] Wang Y, Klijn JGM, Zhang Y, Sieuwerts AM, Look MP, Yanh F, Talantov D, Timmermans M, Gelder, MEMG, Yu J, Jatkoe T, Berns EMJJ, Atkins D and Foekens JA: Gene-expression Profiles to Predict Distant Metastasis of Lymph-Node-Negative Primary Breast Cancer. *Lancet*, **365**, 671-679. 2005.
- [4] Wollbold J, Huber R, Pohlers D, Koczan D, Guthke R, Kinne RW and Gausmann U: Adapted Boolean Network Models for Extracellular Matrix Formation. *BMC Systems Biology*, **3(77)**, 2009.